



# Improving Tail Flick Analgesic Test by a Custom-Made Restraining Chamber

## ARTICLE INFO

### Article Type

Original Research

### Authors

Hossein Azizi<sup>1\*</sup>

S. Mohammad Ahmadi-Soleimani<sup>2\*</sup>

Yadollah Ranjbar-Slamloo<sup>1</sup>

Saeed Semnani<sup>1</sup>

1. Department of Physiology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran

2. Department of Physiology, School of Paramedical Sciences, Torbat Heydariyeh University of Medical Sciences, Torbat Heydariyeh, Iran

## ABSTRACT

The tail flick test is a common method used to evaluate acute pain in animal models. However, there seems to be a technical limitation to the use of this test, deserving a more detailed consideration. The problem is related to the use of physical restraints during the test, which in turn exposes the animal to stress and could therefore undesirably affect the obtained results. In the present report, a newly designed restraining chamber was used to improve the tail flick test regarding the mentioned limitation. Also, the baseline tail flick latency in animals undergoing the test using the classic restrainers was compared to that in those restrained by the newly designed chamber. The results indicated that the baseline tail flick latency was significantly lower in animals restrained by the newly designed box compared to those restrained by the tube-shaped version, which could be attributed to stress-induced analgesia. In conclusion, it is recommended that researchers use boxes similar to the custom-made box used in this study to prevent stress-induced errors when measuring nociceptive thresholds in rats.

**Keywords:** Tail flick, Restraint, Stress, Analgesia, Rat

### \*Correspondence

Address: \*Corresponding authors:

Department of Physiology, School of Medical Sciences, Tarbiat Modares University, Tehran, Iran. Email: azizih@modares.ac.ir

Department of Physiology, School of Paramedical Sciences, Torbat Heydariyeh University of Medical Sciences, Torbat Heydariyeh, Iran. Email: Ahmadiam1@thums.ac.ir

### Article History

Received: April 17 2021

Accepted: May 11 2021

ePublished: August 20, 2021

## 1. Introduction

Researchers evaluating analgesic effects of drugs often use the tail flick test in animal models, such as rats and mice. Although widely accepted in the literature, this method is associated with a technical limitation related to the use of physical restraints during the experiment [1-4]. Physical restraint is actually necessary to place the animal's tail adjacent to a noxious stimulus, especially in the radiant heat version of the test. For this purpose, researchers generally use conventional hard Plexiglas tubes as restrainer (Fig.1C). This might impose some degree of stress-induced analgesia (SIA) on animals, which in turn could result in overestimation of baseline sensitivity. Previous studies have shown that exposure to acute stress (restraining the animal for a while) results in a significant increase in tail flick latency [5, 6] as well as in plasma level of corticosterone in rats [7]. It has been reported that high levels of plasma corticosteroids are associated with analgesia induced by short-term stress [8-10]. In this study, a simple box recently designed in our laboratory was used in the tail flick test to minimize the mentioned stress.

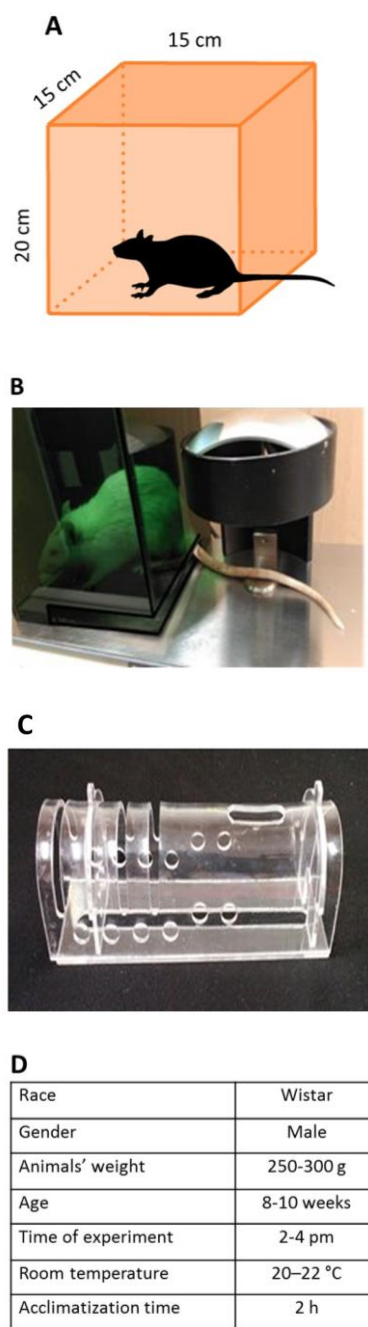
## 2. Methods

As shown in Fig.1A and B, the apparatus used in this study was a rectangular box made of translucent (in this case dark green) Plexiglas with 15×15×20 cm dimensions [11]. In this box, animal's tail could be easily exposed to heat through the gaps located in the inferior parts of the four surrounding dimensions. In contrast to classic tube-shaped restrainers, this box is spacious enough not to expose the animal to physical stress. Thus, the animal is not agitated during the experiment, which seems to be much more ethical from the animal rights viewpoint. Furthermore, it easily gets acclimated to the chamber and its tail could be available for the experimenter. It should be noted that this box could be manually made with a remarkably lower cost than the commercially available restrainers. Therefore, in this study, the baseline tail flick latency was compared between two groups of animals (n=7 per group). Male Wistar rats, weighing 250-300 g, were kept in Plexiglas breeding cages in groups of four animals per cage

with woodchip bedding and free access to food and water. Animals were housed in a colony room at a stable temperature with 12-h light/dark cycles (the light period started at 7 am). All experiments were performed according to the ethical guidelines set by the Ethics Committee of Faculty of Medical Sciences, Tarbiat Modares University based on the NIH Guide for the Care and Use of Laboratory Animals. Animals underwent the test using both types of restrainers (classic tube-shaped ones and the newly designed box). A tail flick analgesiometer apparatus (Harvard, Holliston, US) was used for the test. A light beam, generated by a 150 W electrical bulb, was focused on the dorsal part of the animal's tail via a concave mirror located above the bulb. The same heat intensity was adjusted for both experimental groups. Also, a 10 s cut-off time was considered to prevent damage to the animal's tail.

## 3. Results and discussion

The obtained results indicated that the baseline tail flick latency was significantly lower in animals restrained by the newly designed box than in those restrained by the tube-shaped version (Fig.2). Thus, it seems that a significant degree of SIA is induced in animals prior to the experiment following the application of tube-shaped restrainers. It has been previously shown that restraining the animal could change the analgesic effect of some drugs [12-15]. Undoubtedly, this could adversely affect the final test results, as observed in the phenomenon under study. Therefore, analgesia might be more sensitively measured with lower baseline latencies in animals not exposed to stress. In particular, in studies with low statistical power (i.e. in cases where the investigator needs to reduce the standard error, and/or the expected effect is small), reducing stress is of great importance. With respect to these concerns, authors found the issue worth reporting. By the way, there are other points that should be considered by researchers in assessing thermal nociceptive thresholds in rats. For example, it is highly recommended to avoid working on female subjects because changes in hormonal profile during the estrous cycle adversely affect pain perception [16-18].



**Fig. 1.** Schematic (A) and real (B) pictures of the newly designed box for the tail flick test compared to classic tube-shaped restrainers (C). Information associated with variables in the tail flick experiment (D).

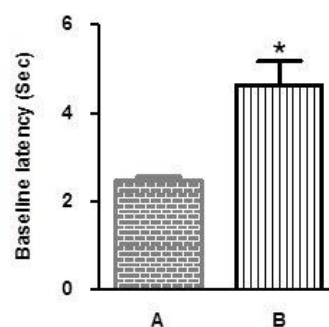
In addition, it is more rational to perform nociceptive tests during adulthood because, for example, pain-modulatory brain structures are fully developed during this period, while during adolescence, the maturation of many regulatory pathways is still in process throughout the nervous system [19]. Finally, researchers are

strongly advised to choose a fixed time for behavioral assessment of nociceptive responses because it has been shown that changes in circadian rhythm potentially affect the results of pain measurement at behavioral level [20].

The new box used in this report could be easily employed in almost all pain studies, especially those assessing the basic mechanisms of pain modulation with an emphasis on nociceptive threshold. Another issue is that researchers currently report changes in pain intensity as normalized values such as the percentage of maximum possible effect (MPE%) [11, 21]. Although this method well indicates the trend of drug effect over time, the final interpretation might be misleading if baseline thresholds are altered by restraint-induced stress. Thus, it is suggested that researchers report changes in baseline thresholds as raw values in their results. This enables the reader to reach a more accurate understanding of basic alterations in cellular responses to painful stimuli.

#### 4. Conclusion

In conclusion, it is recommend that researchers in this field use boxes similar to the custom-made product used in this study to prevent stress-induced errors when measuring nociceptive thresholds in rats.



**Fig. 2.** Comparison of the baseline tail flick latency between the two groups of animals: A) restrained by the newly designed box and B) restrained by a commercially available tube-shaped restrainer. Data are expressed as mean  $\pm$  SEM, \* $p < .05$ ,  $n=7$  per group.

## Acknowledgement

This work was supported by Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran.

## References

1. Mao, J., D.D. Price, and D.J. Mayer, *Thermal hyperalgesia in association with the development of morphine tolerance in rats: roles of excitatory amino acid receptors and protein kinase C*. Journal of Neuroscience, 1994. **14**(4): p. 2301-2312.
2. Singh, P., et al., *Comparison of electroencephalographic changes in response to acute electrical and thermal stimuli with the tail flick and hot plate test in rats administered with opiorphin*. BMC neurology, 2018. **18**(1): p. 1-10.
3. Foroud, M. and N. Vesal. *Evaluation of the anti-nociceptive effects of morphine, tramadol, meloxicam and their combinations using the tail-flick test in rats*. in *Veterinary Research Forum*. 2015. Faculty of Veterinary Medicine, Urmia University, Urmia, Iran.
4. d'Amore, A., F. Chiarotti, and P. Renzi, *High-intensity nociceptive stimuli minimize behavioral effects induced by restraining stress during the tail-flick test*. Journal of pharmacological and toxicological methods, 1992. **27**(4): p. 197-201.
5. Gamaro, G., et al., *The effects of acute and repeated restraint stress on the nociceptive response in rats*. Physiology & behavior, 1998. **63**(4): p. 693-697.
6. Miller, D.B., *Restraint-induced analgesia in the CD-1 mouse: interactions with morphine and time of day*. Brain research, 1988. **473**(2): p. 327-335.
7. Gameiro, G.H., et al., *The effects of restraint stress on nociceptive responses induced by formalin injected in rat's TMJ*. Pharmacology Biochemistry and Behavior, 2005. **82**(2): p. 338-344.
8. Bogdanov, A., et al., *Stress-induced analgesia. The role of corticosteroids*. Fiziologicheskii zhurnal imeni IM Sechenova, 1995. **81**(3): p. 61-67.
9. Sutton, L., et al., *A permissive role of corticosterone in an opioid form of stress-induced analgesia: blockade of opiate analgesia is not due to stress-induced hormone release*. Brain research, 1994. **663**(1): p. 19-29.
10. Yamamoto, A., et al., *Pharmacological relationship between nicotinic and opioid systems in analgesia and corticosterone elevation*. Life sciences, 2011. **89**(25-26): p. 956-961.
11. Ranjbar-Slamloo, Y., et al., *Orexin receptor type-1 antagonist SB-334867 inhibits the development of morphine analgesic tolerance in rats*. Peptides, 2012. **35**(1): p. 56-59.
12. Calcagnetti, D.J. and S.G. Holtzman, *Potentiation of morphine analgesia in rats given a single exposure to restraint stress immobilization*. Pharmacology Biochemistry and Behavior, 1992. **41**(2): p. 449-453.
13. Calcagnetti, D.J., J.L. Stafinsky, and T. Crisp, *A single restraint stress exposure potentiates analgesia induced by intrathecally administered DAGO*. Brain research, 1992. **592**(1-2): p. 305-309.
14. Appelbaum, B.D. and S.G. Holtzman, *Stress-induced changes in the analgesic and thermic effects of opioid peptides in the rat*. Brain research, 1986. **377**(2): p. 330-336.
15. Calcagnetti, D.J. and S.G. Holtzman, *Factors affecting restraint stress-induced potentiation of morphine analgesia*. Brain research, 1990. **537**(1-2): p. 157-162.
16. Frye, C., C. Cuevas, and R. Kanarek, *Diet and estrous cycle influence pain sensitivity in rats*. Pharmacology Biochemistry and Behavior, 1993. **45**(1): p. 255-260.
17. Martínez-Gómez, M., et al., *Assessing pain threshold in the rat: changes with estrus and time of day*. Physiology & behavior, 1994. **55**(4): p. 651-657.
18. Vinogradova, E., D. Zhukov, and A. Batuev, *The effects of stages of the estrous cycle on pain thresholds in female white rats*. Neuroscience and behavioral physiology, 2003. **33**(3): p. 269-272.
19. Salmanzadeh, H., et al., *Adolescent drug exposure: A review of evidence for the development of persistent changes in brain function*. Brain research bulletin, 2020. **156**: p. 105-117.
20. Rosenfeld, J.P. and P.E. Rice, *Diurnal rhythms in nociceptive thresholds of rats*. Physiology & behavior, 1979.
21. Soleimani, S.M.A., et al., *Orexin type 1 receptor antagonism in rat locus coeruleus prevents the analgesic effect of intra-LC met-enkephalin microinjection*. Pharmacology Biochemistry and Behavior, 2015. **136**: p. 102-106.