



Investigating the Potential of Curcumin in Reducing Liver Fibrosis via Inhibition of TGF- β 1 Gene Expression

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ABSTRACT

Introduction: Transforming growth factor (TGF)- β 1 is crucial in developing liver fibrosis. Curcumin has been shown to effectively halt the advancement of liver fibrosis by inhibiting the TGF- β 1/Smad signaling pathway. Nevertheless, curcumin's impact on liver fibrosis regression remains unclear. This study explored the involvement of curcumin and TGF- β 1 in the regression of liver fibrosis.

Methods: An experimental male C57BL/6 mice model included 6 treatment groups. The treatment groups were injected with carbon tetrachloride (CCl₄) for 4 and 6 weeks to induce liver fibrosis, negative controls were injected with olive oil. After cessation of injection, 2 of the treatment groups were given curcumin for 2 weeks. TGF- β 1 expression in liver cells was analyzed by real-time PCR assay. ELISA analyzed hydroxyproline liver tissue levels. Values of $p < 0.05$ were regarded as statistically significant.

Results: CCl₄ injection induced liver fibrosis and significantly increased TGF- β 1 expression and hydroxyproline levels in tissues. Curcumin administration decreased the expression of TGF- β 1 and hydroxyproline levels in the liver and accelerated the regression of liver fibrosis.

Conclusion: Curcumin accelerates regression of liver fibrosis, likely through decreasing TGF- β 1 expression in the liver.

Keywords:

Liver fibrosis, Carbon tetrachloride, Curcumin, TGF- β 1, Hydroxyproline

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INTRODUCTION

Liver diseases account for approximately 2 million deaths annually worldwide, with 1 million cases due to complications of cirrhosis and 1 million cases due to viral hepatitis and hepatocellular carcinoma [1]. The most common causes of chronic liver diseases include hepatitis B virus, hepatitis C virus, alcohol-related liver disease (ALD), and non-alcoholic fatty liver disease (NAFLD) [2]. Liver fibrosis results from a prolonged response in the wound-healing process, triggered by various chronic liver injuries, such as viral infections, autoimmune diseases, metabolic disorders, drug toxicity, alcoholic liver disease, and non-alcoholic fatty liver disease [3]. No matter the cause, clinical evidence indicates that liver fibrosis can regress once the underlying cause is eliminated [4]. The signaling pathways of Transforming Growth Factor Beta1 (TGF- β 1) play a critical role in regulating several cellular processes such as

proliferation, differentiation, migration, and apoptosis, all of which are vital for maintaining balance in tissues and organs. TGF- β 1 is a significant regulator of liver health, influencing all stages of disease development, from the initial liver injury through inflammation and fibrosis to cirrhosis and hepatocellular carcinoma [5]. The pro-fibrogenic effects of TGF- β 1 in liver fibrosis primarily occur through the activation of hepatic stellate cells, which are the main producers of extracellular matrix during liver fibrosis. TGF- β 1 triggers the activation, proliferation, and differentiation of HSCs into myofibroblasts while inhibiting their apoptosis. This leads to excessive synthesis of ECM proteins, including fibronectin and collagens types I, III, and IV. Additionally, TGF- β 1 disrupts the production of matrix-degrading proteases and increases the levels of protease inhibitors, such as tissue inhibitors of metalloproteinases (TIMP) and plasminogen activator inhibitors [6, 7]. Therefore, inhibiting

the TGF- β 1/Smad signaling pathway is crucial for restraining the activation and proliferation of HSCs and improving liver fibrosis [8].

The primary curcuminoid found in turmeric is called curcumin, which is a polyphenol that was first isolated from the rhizome of *Curcuma longa* about two centuries ago. Its structure, known as diferuloylmethane, was identified in 1910 [9]. Curcumin has been extensively studied as an anti-fibrosis agent associated with TGF- β 1. Additionally, several studies using animal models of chronic liver diseases have demonstrated that curcumin exhibits anti-fibrogenic properties by modulating various processes involved in liver damage; This includes targeting different cellular signaling pathways, such as TGF- β 1 and Inhibition of hepatic stellate cell activation, cytokines, transcription factors, and genes regulating cell proliferation and apoptosis [10, 11].

We have consequently assessed the effectiveness of curcumin in treating liver fibrosis induced by CCl₄ in animal models and examined the levels of TGF- β 1 expression and hydroxyproline within this model.

MATERIALS AND METHODS

Experimental Animals

This research was conducted as an in vivo experimental study employing male C57BL/6 mice. A total of 30 male C57BL/6 mice, aged 6-8

weeks and weighing an average of 20 ± 2 g, were utilized in the study. The research was approved and registered by the Ethics Committee of Tarbiat Modarres University of Medical Sciences under a specific code IR.MODARES.AEC.1403.003.

Animal Grouping

1. Control Groups (4 and 6 Weeks): Each group consisted of five mice. These mice received olive oil twice a week for either 4 or 6 weeks.
2. Fibrosis Groups (4 and 6 Weeks): Each group comprised five mice. The mice were treated with carbon tetrachloride (CCl₄) diluted in olive oil at a concentration of 0.5 ml/g BW, administered intraperitoneally.
3. Curcumin Treatment Groups: Each group included five mice with fibrosis. After cessation of CCl₄ injection, These mice were given curcumin(8203540002, Sigma-Aldrich Chemie GmbH, Germany) at a dosage of 100 mg/kg BW every other day for two weeks via gavage.

RNA isolation and real-time PCR

The total RNA was extracted from the mouse liver using TRIzol. Subsequently, cDNA was synthesized using a cDNA Synthesis Kit (A101161, parstous, Iran). Real-time PCR was performed to TGF gene expression (normalized to GAPDH) using specific primers (Table 1) and SYBR Green Master Mix.

Table1. Primers applied for Real-time PCR assays

| Target gene | Forward primer (5'-3') | Reverse primer (5'-3') |
|----------------|-------------------------|------------------------|
| TGF- β 1 | CAACGCCATCTATGAGAAAACCA | AAAGCCCTGTATTCCGTCTCC |
| GAPDH | GAGAGTGTTTCCTCGTCCCGTA | TGCCGTGAGTGAGTCATACT |

ELISA

The concentration of hydroxyproline in liver tissue was measured using a kit(KHPO96, Kiazist, Iran) Tissue samples were homogenized and then digested with a strong acid before oxidizing. The resulting samples were then reacted with a chromogen. The optical density (OD) of each well was measured at a wavelength of 560 nm. The OD values are directly proportional to the collagen concentration in the samples.

Histopathological Evaluation

The liver tissues were fixed in 10% formalin for 24 hours, dehydrated using an alcohol series from 70% to 100% alcohol, and embedded in

paraffin. Thin sections measuring 4-5 micrometers were then cut using a microtome and mounted on glass slides. These sections were stained with Hematoxylin to visualize nuclei and Eosin to highlight cytoplasmic and extracellular matrix components, enabling examination of tissue structure. Slides were observed under a light microscope after being stained with hematoxylin and eosin (H&E).

Statistical Analysis

The data were analyzed using GraphPad Prism, version 9.5.1 for Windows, a statistical analysis program. A 95% confidence interval ($\alpha=0.05$) was used as a decision threshold for significance. Statistical analyses were performed

using one-way ANOVA followed by Tukey's post hoc test. Results are expressed as mean \pm SD.

RESULT

Results Histopathological analysis

The staining of Hematoxylin and eosin (H&E) showed normal liver architecture in control mice.

Fibrotic mice showed disrupted architecture with cell death and infiltration of mononuclear inflammatory cells around vessels and liver parenchyma. Curcumin-treated mice showed reduced inflammation, cell death, and partial tissue repair

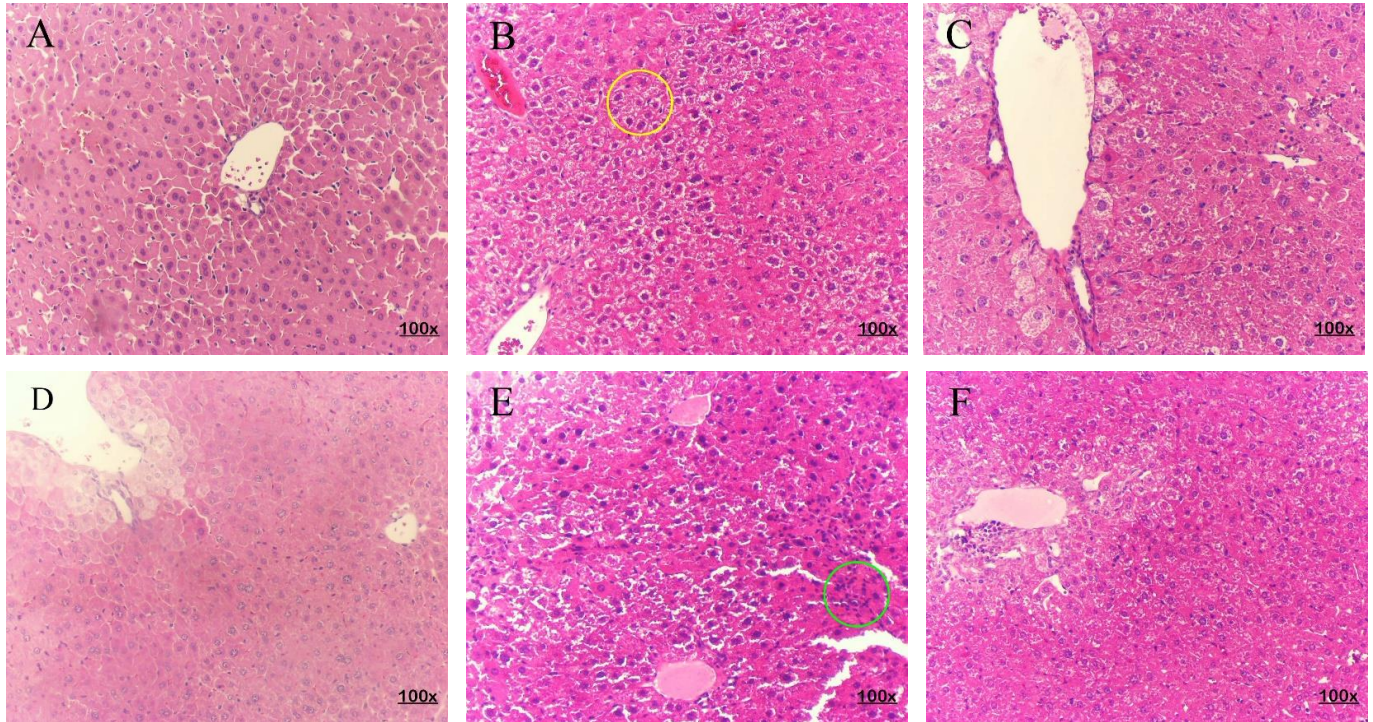


Figure 1. Representative liver histopathology of H&E-stained mice livers in different experimental groups. A: Control, treated with olive oil for 4 weeks; B: Liver fibrosis, treated with CCl₄ for 4 weeks (The circle indicates necrotic cells.); C: Curcumin-treated liver fibrosis, treated with CCl₄ 4 weeks + curcumin 2 weeks; D: Control, treated with olive oil 6 weeks; E: Liver fibrosis, treated with CCl₄ 6 weeks (The circle indicates mononuclear inflammatory cells.); F: Curcumin-treated liver fibrosis, treated with CCl₄ 4 weeks + curcumin 2 weeks.

Measurement of Hydroxyproline Levels in Liver Tissue

Hydroxyproline levels in the 4-week CCl₄-induced liver fibrosis group significantly increased compared to the 4-week control group that received olive oil ($P < 0.01$). In the group with 4-week CCl₄-induced liver fibrosis, followed by two weeks of curcumin treatment, hydroxyproline levels significantly decreased ($P < 0.01$). Similarly, hydroxyproline levels in the 6-week CCl₄-induced liver fibrosis group significantly increased compared to the 6-week control group that received olive oil ($P < 0.001$). In the 6-week CCl₄-induced liver fibrosis group, followed by two weeks of curcumin treatment, hydroxyproline levels also significantly decreased ($P < 0.01$). Additionally, hydroxyproline levels were significantly higher in the 6-week fibrosis group compared to the 4-week fibrosis group ($P < 0.001$).

Although curcumin treatment significantly reduced hydroxyproline levels in the 6-week fibrosis group, they did not return to control levels (Figure 2).

mRNA Expression of the TGF- β 1 Gene

In the 4-week CCl₄-induced liver fibrosis group, TGF- β 1 gene expression significantly increased compared to the 4-week control group that received olive oil ($P < 0.001$). However, following two weeks of curcumin treatment, expression levels significantly decreased ($P < 0.01$). A similar trend was observed in the 6-week CCl₄-induced liver fibrosis group, where gene expression markedly increased compared to the corresponding 6-week control group ($P < 0.0001$), but was significantly reduced after two weeks of curcumin treatment ($P < 0.05$). Furthermore, the TGF- β 1 expression level in the 6-week fibrosis group was notably higher than that in the 4-week fibrosis group ($P < 0.05$) (Figure 3).

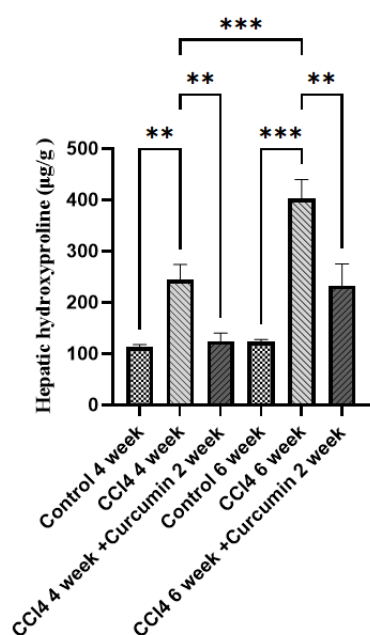


Figure 2: Changes in liver tissue hydroxyproline levels across the study groups. Data are presented as Mean \pm SD. *P-value < 0.05, ** P-value < 0.01, *** P-value < 0.001, **** P-value < 0.0001, compared to the respective control groups.

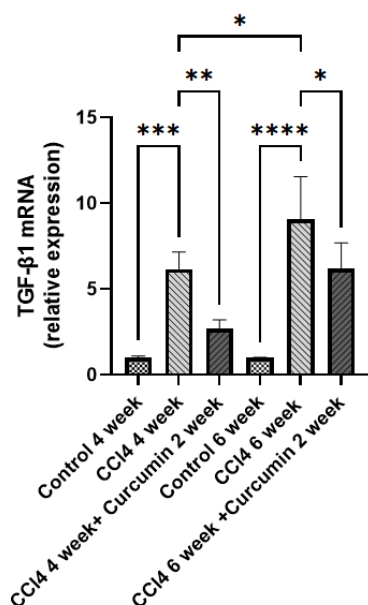


Figure 3: Expression of the TGF-β1 gene in the study groups. Data are presented as mean \pm SD. * P-value < 0.05, ** P-value < 0.01, *** P-value < 0.001, **** P-value < 0.0001.

DISCUSSION

Liver fibrosis is a consequence of various chronic conditions and frequently progresses to cirrhosis. Cirrhosis is linked to severe and life-threatening complications, such as liver failure, portal hypertension, and hepatocellular carcinoma, and is a leading cause of both morbidity and mortality globally.[12, 13] However, clinical and experimental evidence has shown that even advanced fibrosis and cirrhosis

can be reversed [12-15]. Thus, preventing the progression from fibrosis to cirrhosis is regarded as a key goal in the management of patients with liver diseases [16].

This research examined the therapeutic potential of curcumin in treating liver fibrosis after its etiology has been removed using a CCl₄-induced mouse model. Histopathological analysis showed a significant improvement in the liver structure. Also, the level of hydroxyproline and TGF beta 1 gene expression showed a significant decrease in the group treated with curcumin compared to the fibrotic group.

The CCl₄ liver fibrosis model is the most widely used toxic model for inducing liver fibrosis globally. The mechanism of liver fibrosis induced by CCl₄ has been extensively studied, and the most effective administration route is through intraperitoneal injection of CCl₄ [17, 18]. The present study observed that TGF-β1 gene expression and hydroxyproline levels in the 6-week liver fibrosis group were significantly higher than in the 4-week liver fibrosis group. It was a week. This difference may be due to the longer duration of exposure to carbon tetrachloride (CCl₄) in the 6-week liver fibrosis group. As mentioned in previous studies, increasing the dose and duration of CCl₄ exposure aggravated the liver damage and accelerated the progression of fibrosis. Studies have shown that carbon tetrachloride significantly increases the level of fibrotic genes such as TGF-β1 by causing oxidative damage and chronic inflammation in the liver. It causes the accumulation of collagen and fibrosis [17, 19].

Liver fibrosis is characterized by progressive accumulation of extracellular matrix (ECM), which destroys the physiological architecture of the liver [20]. TGF-β1 is found in the liver at all stages of the development of pathological conditions. Early on, it induces apoptosis of hepatocytes and differentiation of stellate cells into myofibroblasts after chronic injury, leading to the inflammatory process and liver fibrosis. also acts to promote the proliferation of stellate cells and maintain the myofibroblastic phenotype, leading to cirrhosis [21]. TGF-β1 is the key cytokine driving liver fibrogenesis, and inhibiting its synthesis is one of the primary targets in creating antifibrotic drugs [16, 22].

Curcumin has potential utility in the prevention and treatment of various types of liver

diseases, including types of liver diseases, including non-alcoholic and alcoholic fatty liver, fibrosis, cirrhosis, and liver cancer [10].

We have demonstrated that curcumin treatment significantly reduced the expression levels of TGF- β 1 in liver tissue (Figure 3). The reduction in of expression TGF- β 1 was associated with a decrease in the degree of liver fibrosis. Curcumin's pronounced mitigation of CCl₄-induced liver inflammation and fibrosis likely occurred through the inhibition of the TGF- β 1/Smad signaling pathway [23, 24]. In addition, the level of hydroxyproline, which is an indicator of the amount of collagen in the liver tissue, was significantly reduced in the groups treated with curcumin. Our results are consistent with previous findings that showed Curcumin not only inhibits the proliferation of hepatic stellate cells (HSCs) and their transformation into an activated phenotype; but also promotes HSC apoptosis. Inducing apoptosis in HSCs reduces the secretion of type I collagen and decreases TGF- β 1 production [25, 26]. Overall, curcumin may safely improve liver fibrosis by several different mechanisms.

CONCLUSION

our findings demonstrate that the administration of curcumin effectively protected mice livers from CCl₄-induced liver injury and fibrogenesis possibly by reducing TGF- β 1 levels. Further research is needed to explain these mechanisms in more detail.

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CONFLICTS OF INTEREST

There is no conflict of interest.

REFERENCES

- [1] Asrani SK, Devarbhavi H, Eaton J, Kamath PS. Burden of liver diseases in the world. *Journal of hepatology*. 2019;70(1):151-71.
- [2] Moon AM, Singal AG, Tapper EB. Contemporary Epidemiology of Chronic Liver Disease and Cirrhosis. *Clinical gastroenterology and hepatology : the official clinical practice journal of the American Gastroenterological Association*. 2020;18(12):2650-66.
- [3] Sarin SK, Kumar M, Eslam M, George J, Al Mahtab M, Akbar SMF, et al. Liver diseases in the Asia-Pacific region: a Lancet Gastroenterology & Hepatology Commission. *The lancet Gastroenterology & hepatology*. 2020;5(2):167-228.
- [4] Ellis EL, Mann DA. Clinical evidence for the regression of liver fibrosis. *Journal of hepatology*. 2012;56(5):1171-80.
- [5] Dooley S, ten Dijke P. TGF- β in progression of liver disease. *Cell and tissue research*. 2012;347(1):245-56.
- [6] Dewidar B, Meyer C, Dooley S, Meindl-Beinker AN. TGF- β in Hepatic Stellate Cell Activation and Liver Fibrogenesis-Updated 2019. *Cells*. 2019;8(11).
- [7] Xu F, Liu C, Zhou D, Zhang L. TGF- β /SMAD Pathway and Its Regulation in Hepatic Fibrosis. *The journal of histochemistry and cytochemistry : official journal of the Histochemistry Society*. 2016;64(3):157-67.
- [8] Tan Z, Sun H, Xue T, Gan C, Liu H, Xie Y, et al. Liver Fibrosis: Therapeutic Targets and Advances in Drug Therapy. *Frontiers in cell and developmental biology*. 2021;9:730176.
- [9] Aggarwal BB, Sundaram C, Malani N, Ichikawa H. Curcumin: the Indian solid gold. *Advances in experimental medicine and biology*. 2007;595:1-75.
- [10] Vera-Ramirez L, Pérez-Lopez P, Varela-Lopez A, Ramirez-Tortosa M, Battino M, Quiles JL. Curcumin and liver disease. *BioFactors* (Oxford, England). 2013;39(1):88-100.
- [11] Wu P, Huang R, Xiong YL, Wu C. Protective effects of curcumin against liver fibrosis through modulating DNA methylation. *Chinese journal of natural medicines*. 2016;14(4):255-64.
- [12] Friedman SL. Mechanisms of hepatic fibrogenesis. *Gastroenterology*. 2008;134(6):1655-69.
- [13] Bataller R, Brenner DA. Liver fibrosis. *The Journal of clinical investigation*. 2005;115(2):209-18.
- [14] Iredale JP. Models of liver fibrosis:

- exploring the dynamic nature of inflammation and repair in a solid organ. *The Journal of clinical investigation*. 2007;117(3):539-48.
- [15] Lee UE, Friedman SL. Mechanisms of hepatic fibrogenesis. *Best practice & research Clinical gastroenterology*. 2011;25(2):195-206.
- [16] Friedman SL. Evolving challenges in hepatic fibrosis. *Nature reviews Gastroenterology & hepatology*. 2010;7(8):425-36.
- [17] Scholten D, Trebicka J, Liedtke C, Weiskirchen R. The carbon tetrachloride model in mice. *Laboratory animals*. 2015;49(1 Suppl):4-11.
- [18] Boll M, Weber LW, Becker E, Stampfl A. Mechanism of carbon tetrachloride-induced hepatotoxicity. Hepatocellular damage by reactive carbon tetrachloride metabolites. *Zeitschrift fur Naturforschung C, Journal of biosciences*. 2001;56(7-8):649-59.
- [19] Fujii T, Fuchs BC, Yamada S, Lauwers GY, Kulu Y, Goodwin JM, et al. Mouse model of carbon tetrachloride induced liver fibrosis: Histopathological changes and expression of CD133 and epidermal growth factor. *BMC gastroenterology*. 2010;10:79.
- [20] Roehlen N, Crouchet E, Baumert TF. Liver Fibrosis: Mechanistic Concepts and Therapeutic Perspectives. *Cells*. 2020;9(4).
- [21] Fabregat I, Moreno-Càceres J, Sánchez A, Dooley S, Dewidar B, Giannelli G, et al. TGF- β signalling and liver disease. *The FEBS journal*. 2016;283(12):2219-32.
- [22] Gressner AM, Weiskirchen R. Modern pathogenetic concepts of liver fibrosis suggest stellate cells and TGF-beta as major players and therapeutic targets. *Journal of cellular and molecular medicine*. 2006;10(1):76-99.
- [23] Yao Q-y, Xu B-l, Wang J-y, Liu H-c, Zhang S-c, Tu C-t. Inhibition by curcumin of multiple sites of the transforming growth factor-beta1 signalling pathway ameliorates the progression of liver fibrosis induced by carbon tetrachloride in rats. *BMC Complementary and Alternative Medicine*. 2012;12(1):156.
- [24] Hernández-Aquino E, Quezada-Ramírez MA, Silva-Olivares A, Ramos-Tovar E, Flores-Beltrán RE, Segovia J, et al. Curcumin downregulates Smad pathways and reduces hepatic stellate cells activation in experimental fibrosis. *Annals of hepatology*. 2020;19(5):497-506.
- [25] Kang HC, Nan JX, Park PH, Kim JY, Lee SH, Woo SW, et al. Curcumin inhibits collagen synthesis and hepatic stellate cell activation in-vivo and in-vitro. *The Journal of pharmacy and pharmacology*. 2002;54(1):119-26.
- [26] Shu JC, He YJ, Lv X, Zhao JR, Zhao J, Shen Y, et al. Effect of curcumin on the proliferation and apoptosis of hepatic stellate cells. *Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas*. 2009;42(12):1173-8.